



Analysis of Microbial Load in Air at Different Time Interval in Dental Clinic

S. Saivarshine ^a and N. P. Muralidharan ^{b*}

^a *Saveetha Dental College and Hospitals, Saveetha Institute of Medical and Technical Sciences, Saveetha University, Chennai, Tamil Nadu, India.*

^b *Department of Microbiology, Saveetha Dental College and Hospitals, Saveetha Institute of Medical and Technical Sciences (SIMATS), Saveetha University Chennai, Tamil Nadu, India.*

Authors' contributions

This work was carried out in collaboration between both authors. Both authors read and approved the final manuscript.

Article Information

DOI: 10.9734/JPRI/2021/v33i62A35670

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/74346>

Original Research Article

Received 22 October 2021
Accepted 27 December 2021
Published 28 December 2021

ABSTRACT

Introduction: Patients and healthcare professionals can be exposed to several microorganisms that colonize or invade the oral cavity and respiratory tract, or are transported in the water used during treatment, which increases the risk of infection.

Materials and Methods: To determine the air germ count over four consecutive hours, by exposed plate method using Brain heart infusion agar (BHI) plates in the multi chair treatment room of the Department of Prosthodontics, Department of Periodontics, Department of Pedodontics, Department of Conservative dentistry & Operative Dentistry of Saveetha Dental College.

Results: The CFUs in the multi-chair treatment room were between 47 and 243 CFU m³. . During treatment, it reached up to 243 CFU m³.

Conclusion: During treatment, the bacterial count was greater than the actual time before treatment. While bacterial numbers in dental rooms have been substantially higher, the risk in dental clinics is higher due to the formation of aerosols that contain microorganisms.

Keywords: *Air; dental clinic; microbial load; time duration.*

[≡] Graduate,

[°] Assistant professor,

*Corresponding author: E-mail: muralidharan@saveetha.com;

1. INTRODUCTION

Patients and healthcare professionals can be exposed to several microorganisms that colonize or invade the oral cavity and respiratory tract, or are transported in the water used during treatment, which increases the risk of infection [1-3].

Water lines, microbial aerosols, and clinical contact surfaces are three of the most common causes of infection. Microorganisms that multiply on the inner surface of water tubes and form a biofilm may colonize dental unit waterlines [4-6]. The biofilm will then offer some protection to maximize the number of microorganisms in the water used for dental treatments. Microbial aerosols are frequently generated during dental procedures, smaller particles can float in the air and enter small passages of the lungs, whereas larger particles can easily settle on environmental surfaces [7-10].

Some surfaces, particularly so-called clinical contact surfaces that are constantly handled (e.g. dental unit switches, lamp handles, and drawer knobs), can serve as microorganism reservoirs.

Microorganisms can be transmitted to instruments, other environmental objects, or the nose, mouth, or eyes of healthcare workers and patients as these surfaces are touched. While it has been recognized that environmental matrices (water, air, and clinical touch surfaces) can play an important role as a vector for infection, the research on microbial pollution in the dental clinic setting is not comprehensive. Existing trials included only a small number of hospitals and very few measured total microbial environmental degradation. Moreover, while the assessment of water toxicity is based on widely agreed and uniform sampling and processing procedures and well-specified threshold values [11-21].

For this purpose, hospital environmental management protocols may provide useful assistance for the reduction of nosocomial infections [22,23]. This is especially the case in high-risk health services where people are more vulnerable due to their health problems or in operating theatres due to proximity to air tissue [24,25]. In reality, surgeons were the first to work with environmental hygiene conditions during high-risk surgery to eliminate post-operative

infections. Since then, several authors have emphasized the value of microbial monitoring of environmental matrices.

Mick et al. [26] first observed aerosol particles produced during dental treatment. The researchers have established their new fields of study in dental aerobiology. Besides, the authors characterized their studies as a science of air particles and the relationship between these particles and the wellbeing of patients and the treatment of staff. They also found that streptococcal aerosols emitted by dental turbines remained detectable in the air after 24 hours. However, improvements in dental chair technology and the use of rubber dams should also be taken into account when considering the possibility of dental aerosols.

While dental therapies have been shown to greatly increase the amount of bacterial air pollution, there is currently no systematic study of the microbiological atmosphere in dental practice and a multi-stage dental clinic compared to public areas. In comparison, the experiments carried out concerned a few assessment points before and during dental procedures. To date, data over longer periods of several hours and days have not been produced.

This study aimed to analyze the airborne microbial load at a normal dental practice and period of four consecutive hours.

2. MATERIALS AND METHODS

To determine the aerosol microorganism by exposed plate method over four consecutive hours, on brain heart infusion agar (BHI) plates in the multi chair treatment room of the Department of Prosthodontics, Department of Periodontics, Department of Pedodontics, Department of Conservative dentistry & Operative Dentistry of Saveetha Dental College. Particles, bacteria, and fungi were deposited on the culture medium located underneath. Windows and doors were kept closed during the day. In the multi-chair dental clinic as well as in the dental practice complex dental treatments such as professional tooth cleaning, restoration, dental fillings, and root canal treatments were performed during the air sampling. During the dental treatment, the windows were closed and the clinics were using air conditioning units.

3. RESULTS



Fig. 1. Culture plates collected and incubated (24-48hrs) between 9 am-10 am from different clinics



Fig. 2. Culture plates collected and incubated (24-48hrs) between 10 am-11 am from different clinics

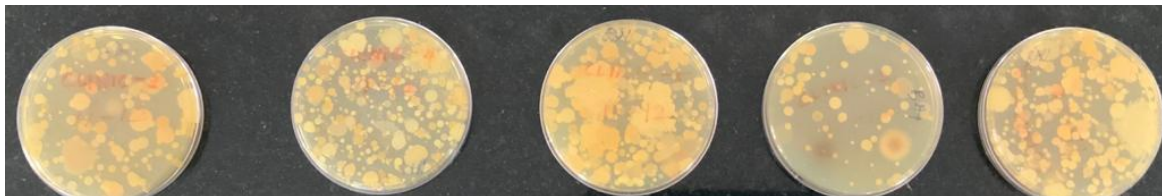


Fig. 3. Culture plates collected and incubated (24-48hrs) between 11 am-12 pm from different clinics

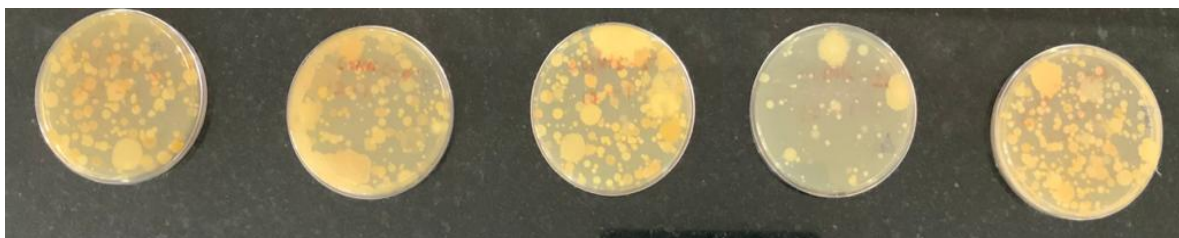


Fig. 4. Culture plates collected and incubated (24-48hrs) between 12 pm-1 pm from different Clinics

Table 1. The table depicts the Colony forming units (CFUs) between consecutive time intervals in different clinics

CLINIC	9AM-10AM	10AM-11AM	11AM-12 PM	12PM-1PM
CLINIC-1	167	115	124	107
CLINIC-2	243	135	235	144
CLINIC-3	151	146	199	155
CLINIC-4	96	138	47	117
CLINIC-5	216	116	183	190

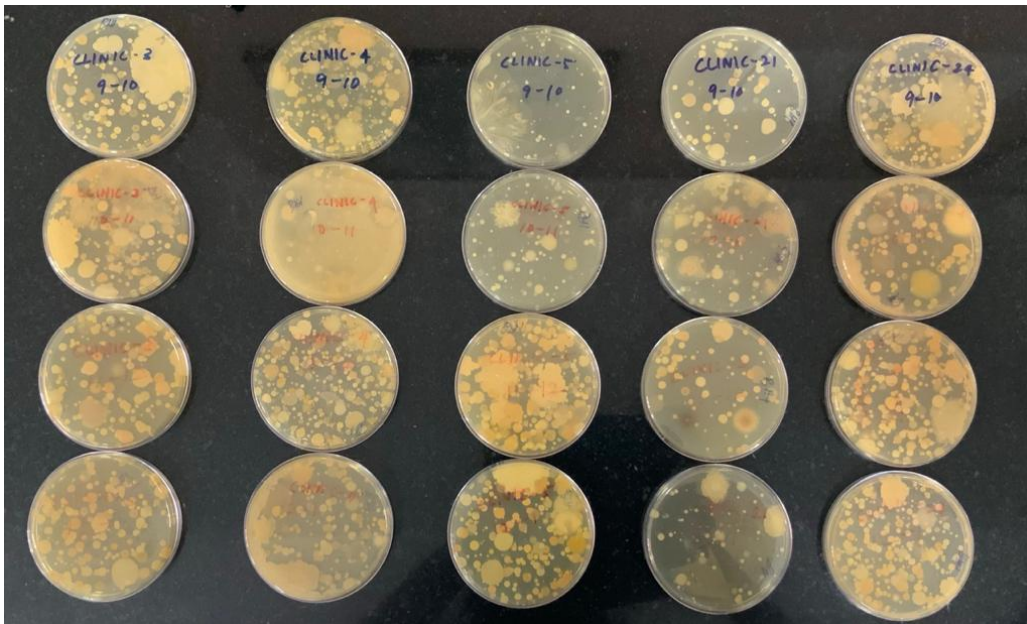


Fig. 5. Culture plates collected and incubated (24-48hrs) from different clinics between 9am-10am, 10am-11am, 11am-12pm and 12pm-1pm

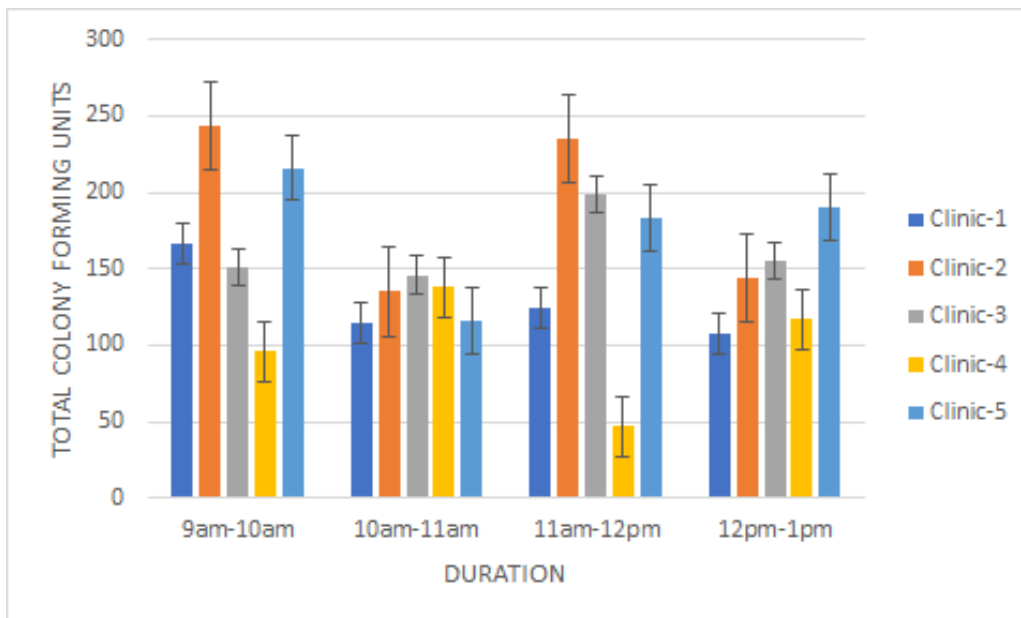


Fig. 6. The graph represents the no. of colony-forming units (CFUs) formed in the clinics between the duration of 9 am-10 am, 10 am-11 am, 11 am-12 pm, and 12 pm-1 pm. The X-axis represents duration and the Y-axis represents the total number of colony-forming units

4. DISCUSSION

4.1 Total airborne Microorganisms

Fig. 1. shows the culture plates collected and incubated (24-48hrs) between 9 am-10 am from different clinics. The total colony-forming units

(CFUs) in the clinics during this hour, Clinic-1, Clinic-2, Clinic-3, Clinic-4, and Clinic-5 with 167 CFUs, 243 CFUs, 151 CFUs, 96 CFUs, and 216 CFUs respectively. A maximum of 243 CFUs was identified in Clinic-2 and a minimum of 96 CFUs was identified in Clinic-4. Fig. 2. shows the culture plates collected and incubated (24-48hrs) between 10 am-11 am from different clinics. The

total colony-forming units (CFUs) in the clinics during this hour, Clinic-1, Clinic-2, Clinic-3, Clinic-4, and Clinic-5 with 115 CFUs, 135 CFUs, 146 CFUs, 138 CFUs, and 116 CFUs respectively. A maximum of 146 CFUs was identified in Clinic-3 and a minimum of 115 CFUs was identified in Clinic-1. Fig. 3. shows the culture plates collected and incubated (24-48hrs) between 11 am-12 pm from different clinics. The total colony-forming units (CFUs) in the clinics during this hour, Clinic-1, Clinic-2, Clinic-3, Clinic-4, and Clinic-5 with 124 CFUs, 235 CFUs, 199 CFUs, 47 CFUs, and 183 CFUs respectively. A maximum of 235 CFUs was identified in Clinic-2 and a minimum of 47 CFUs was identified in Clinic-4. Fig. 4. shows the culture plates collected and incubated (24-48hrs) between 12 pm - 1 pm from different clinics. The total colony-forming units (CFUs) in the clinics during this hour, Clinic-1, Clinic-2, Clinic-3, Clinic-4, and Clinic-5 with 107 CFUs, 144 CFUs, 155 CFUs, 117 CFUs, and 190 CFUs respectively. A maximum of 190 CFUs was identified in Clinic-5 and a minimum of 107 CFUs was identified in Clinic-1.

Microbial aerosols in the dental clinic may have various causes, e.g. from dental procedures, dental staff, or patients, but also outside sources, i.e. air, soil, and dust. Such aerosols can transmit microorganisms to dental staff or patients. Due to varying potential causes of microbial air pollution, quantitative and qualitative studies of airborne microbes in dental clinics compared to the public sector may be useful in estimating the risk of infection due to microbial aerosols throughout dental surgery. This has not been available in literature so far. To date, there is no comparison of microbiological evidence from aerosols obtained using the same methodology and derived from dental and communal settings. Different air sampling would yield different results, making it much more impossible to compare different dental procedures. This makes it impossible to estimate the effect of microbial aerosols on dental work.

The estimation of the overall germ count at various time points in the day facilitates a detailed analysis of the shifts in the microbial load. The variations in the experimental architectures and the technological designs of the air sampling systems have often had to be taken into account when comparing the studies carried out so far. Also, the time intervals observed were often inconsistent between these earlier studies. The values detected by Castiglia

[15] were 107 CFU m³ on average (baseline) and there was no quantitative discrepancy between the empty room values and those detected during the presence of patients and the treatment team. As a result, the investigators found that the presence of humans did not result in a substantial rise in air germ values. This is consistent with the findings given by Castiglia [15]. The sampling position chosen in the room should not be too close to the patient. Particles with a diameter between 50 mm and 100 mm display ballistic activity if the forces of inertia are stronger than the forces of friction. Based on their composition, a contorted direction of motion similar to that of a projectile is taken and, after a brief time in the air, the particles adsorb onto the neighboring surface [27]. Particles less than 50 mm are undetectable to the human eye but can stay in the air as aerosols for a long period. In this study, the assessment of pure airborne microbes in aerosol particles is not affected by direct spraying because the measurement was not carried out too close to the treatment chair. Compared to this report, Bennett et al. [14] observed comparable CFU-values for the most part in general dental practice, with average values of approximately 500 CFU m³. However, these values have consistently been surpassed by considerably higher peaks of up to 6000 CFU m³. Szymanska and Dutkiewicz [28] received maximum values of up to 40,080 CFU m³. The risk of external pollution must be considered for such high values. The act of suction of between 100 and 600 of air has demonstrated its robustness, given its capacity to be assessed and the knowledge value obtained. Smaller concentrations of the air are too inaccurate and poor in microbial counts because too few microbes have been captured. Excessively long period measurements are less than suitable for the representation of shifts because the points in time with higher loads are marginally leveled and obliterated by the use of a single average value, but which cover periods overlapping various dental procedures or events. However, the measurements were taken by Grenier [29] over a time of 30 minutes. The airflow was just 20 l/min. Very few experiments have been conducted to date with such depth that various forms of microbes are found in dental aerosols relative to airborne microbes from other non-dental public areas using limited agar or biochemical norm measures. Al Maghlouth et al. [12] recorded a CONS proportion of 37.7 percent and a micrococci proportion of 32.6 percent. This is consistent with the findings of the present analysis. The pseudomonas bacteria accounted

for 0.6 percent and the fungi accounted for 0.9 percent of the microbes measured. Pagniano [30] examined 166 microbes randomly collected using normal methods and observed comparable amounts. It really should be emphasized that the reduced capacity of the API1 test and related systems present a challenge specifically in the classification of non-pathogenic environmental microbes. These devices have been designed for the rapid detection of some clinically important bacteria and are thus the only species that can be identified. This study analyzed the microbial load in the air at different intervals in the dental clinic.

Our team has extensive knowledge and research experience that has translated into high-quality publications [31-50].

5. CONCLUSION

During treatment, the bacterial count was greater than the actual time before treatment. While bacterial numbers in dental rooms have been substantially higher, the risk from dental clinics is higher in the microorganisms, host susceptibility, and exposure period.

CONSENT

It is not applicable.

ETHICAL APPROVAL

It is not applicable.

ACKNOWLEDGEMENT

We would like to thank Saveetha Dental College for supporting us to conduct the study.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Guidelines for Infection Control in Dental Health-Care Settings. [Internet]. Psyc EXTRA Dataset; 2003. Available: <http://dx.doi.org/10.1037/e545872006-001>
2. Harrel SK, Molinari J. Aerosols and splatter in dentistry [Internet]. The Journal of the American Dental Association. 2004;135:429–37. Available: <http://dx.doi.org/10.14219/jada.archive.2004.0207>
3. Leggat PA, Kedjarune U. Bacterial aerosols in the dental clinic: a review [Internet]. International Dental Journal. 2001;51:39–44. Available: <http://dx.doi.org/10.1002/j.1875-595x.2001.tb00816.x>
4. Barbeau J, Tanguay R, Faucher E, Avezard C, Trudel L, Côté L, et al. Multiparametric analysis of waterline contamination in dental units. Appl Environ Microbiol. 1996;62(11):3954–9.
5. Coleman DC, O'Donnell MJ, Shore AC, Russell RJ. Biofilm problems in dental unit water systems and its practical control [Internet]. Journal of Applied Microbiology. 2009;106:1424–37. Available: <http://dx.doi.org/10.1111/j.1365-2672.2008.04100.x>
6. Porteous N, Sun Y, Dang S, Schoolfield J. A comparison of 2 laboratory methods to test dental unit waterline water quality [Internet]. Diagnostic Microbiology and Infectious Disease. 2013;77:206–8. Available: <http://dx.doi.org/10.1016/j.diagmicrobio.2013.07.010>
7. Decraene V, Ready D, Pratten J, Wilson M. Air-borne microbial contamination of surfaces in a UK dental clinic [Internet]. The Journal of General and Applied Microbiology. 2008;54:195–203. Available: <http://dx.doi.org/10.2323/jgam.54.195>
8. Prospero E, Savini S, Annino I. Microbial Aerosol Contamination of Dental Healthcare Workers' Faces and Other Surfaces in Dental Practice [Internet]. Infection Control & Hospital Epidemiology. 2003;24:139–41. Available: <http://dx.doi.org/10.1086/502172>
9. Rautemaa R, Nordberg A, Wuolijoki-Saaristo K, Meurman JH. Bacterial aerosols in dental practice – a potential hospital infection problem? [Internet]. Journal of Hospital Infection. 2006;64:76–81. Available: <http://dx.doi.org/10.1016/j.jhin.2006.04.011>
10. Timmerman MF, Menso L, Steinfors J, van Winkelhoff AJ, van der Weijden GA. Atmospheric contamination during ultrasonic scaling [Internet]. Journal of Clinical Periodontology. 2004;31:458–62.

- Available:<http://dx.doi.org/10.1111/j.1600-051x.2004.00511.x>
11. Acharya S, Priya H, Purohit B, Bhat M. Aerosol contamination in a rural university dental clinic in south India [Internet]. *International Journal of Infection Control*. 2010;6. Available:<http://dx.doi.org/10.3396/ijic.v6i1.003.10>
 12. Maghlouth AA, Al Maghlouth A, Al Yousef Y, Al Bagieh N. Qualitative and Quantitative Analysis of Bacterial Aerosols [Internet]. *The Journal of Contemporary Dental Practice*. 2004;5:91–100. Available:<http://dx.doi.org/10.5005/jcdp-5-4-91>
 13. Angelillo IF, D'Errico MM, Pavia M, Prospero E, Romano F. [Evaluation of microbial air contamination in dental areas]. *Arch Stomatol*. 1990;31(3):511–8.
 14. Bennett AM, Fulford MR, Walker JT, Bradshaw DJ, Martin MV, Marsh PD. Microbial aerosols in general dental practice [Internet]. *British Dental Journal*. 2000;189:664–7. Available:<http://dx.doi.org/10.1038/sj.bdj.4800859>
 15. Castiglia P, SItI Working Group Hygiene in Dentistry, Liguori G, Montagna MT, Napoli C, Pasquarella C, et al. Italian multicenter study on infection hazards during dental practice: Control of environmental microbial contamination in public dental surgeries [Internet]. *BMC Public Health*. 2008;8. Available:<http://dx.doi.org/10.1186/1471-2458-8-187>
 16. Cellini L. Quantitative microbial monitoring in a dental office [Internet]. *Public Health*. 2001;115:301–5. Available: [http://dx.doi.org/10.1016/s0033-3506\(01\)00464-4](http://dx.doi.org/10.1016/s0033-3506(01)00464-4)
 17. Cristina ML, Spagnolo AM, Sartini M, Dallera M, Ottria G, Lombardi R, et al. Evaluation of the risk of infection through exposure to aerosols and spatters in dentistry [Internet]. *American Journal of Infection Control*. 2008;36:304–7. Available:<http://dx.doi.org/10.1016/j.ajic.2007.07.019>
 18. Monarca S. Evaluation of environmental bacterial contamination and procedures to control cross infection in a sample of Italian dental surgeries [Internet]., *Occupational and Environmental Medicine*. 2000;57: 721–6. Available:<http://dx.doi.org/10.1136/oem.57.11.721>
 19. Souza-Gugelmin MCM de, de Souza-Gugelmin MCM, Della Torre Lima C, de Lima SNM, Mian H, Ito IY. Microbial contamination in dental unit waterlines [Internet]. *Brazilian Dental Journal*. 2003;14:55–7. Available:<http://dx.doi.org/10.1590/s0103-64402003000100010>
 20. Singh T, Coogan MM. Isolation of pathogenic Legionella species and legionella-laden amoebae in dental unit waterlines [Internet]. *Journal of Hospital Infection*. 2005;61:257–62. Available:<http://dx.doi.org/10.1016/j.jhin.2005.05.001>
 21. Tanzi ML, Capobianco E, Affanni P, Pizzi S, Vitali P, Veronesi L. Legionella spp. in hospital dental facilities [Internet]. *Journal of Hospital Infection*. 2006;63:232–4. Available:<http://dx.doi.org/10.1016/j.jhin.2006.01.026>
 22. Guidelines for Preventing Health-Care-Associated Pneumonia, 2003: Recommendations of CDC and the Healthcare Infection Control Practices Advisory Committee [Internet]. *Psyc EXTRA Dataset*; 2004. Available:<http://dx.doi.org/10.1037/e548652006-001>
 23. Beldi G, Bisch-Knaden S, Banz V, Mühlemann K, Candinas D. Impact of intraoperative behavior on surgical site infections [Internet]. *The American Journal of Surgery*. 2009;198:157–62. Available:<http://dx.doi.org/10.1016/j.amjsurg.2008.09.023>
 24. Prevention of Surgical-Site Infections [Internet]. *New England Journal of Medicine*. 2010;362:1540–4. Available:<http://dx.doi.org/10.1056/nejmc1002218>
 25. Barrow C. A Patient's Journey through the Operating Department from an Infection Control Perspective [Internet]. *Journal of Perioperative Practice*. 2009;19:94–8. Available:<http://dx.doi.org/10.1177/175045890901900302>
 26. Micik RE, Miller RL, Mazzarella MA, Ryge G. Studies on Dental Aerobiology: I. Bacterial Aerosols Generated during Dental Procedures [Internet]. *Journal of Dental Research*. 1969;48:49–56. Available:<http://dx.doi.org/10.1177/00220345690480012401>

27. Serke M. Sicherer Umgang mit Zytostatika [Internet]. Atemwegs- und Lungenkrankheiten. 2016;42:313–7. Available:<http://dx.doi.org/10.5414/atx02109>
28. Szymańska J, Dutkiewicz J. Concentration and species composition of aerobic and facultatively anaerobic bacteria released to the air of a dental operation area before and after disinfection of dental unit waterlines. *Ann Agric Environ Med*. 2008; 15(2):301–7.
29. Grenier D. Quantitative analysis of bacterial aerosols in two different dental clinic environments. *Appl Environ Microbiol*. 1995;61(8):3165–8.
30. Pagniano RP, Scheid RC, Rosen S, Beck FM. Airborne microorganisms collected in a preclinical dental laboratory [Internet]. *Journal of Dental Education*. 1985;49:653–5. Available: <http://dx.doi.org/10.1002/j.0022-0337.1985.49.9.tb01923.x>
31. Priyadharsini JV, Vijayashree Priyadharsini J, Smiline Girija AS, Paramasivam A. In silico analysis of virulence genes in an emerging dental pathogen *A. baumannii* and related species [Internet]. *Archives of Oral Biology*. 2018;94:93–8. Available:<http://dx.doi.org/10.1016/j.archoralbio.2018.07.001>
32. Vijayashree Priyadharsini J. In silico validation of the non-antibiotic drugs acetaminophen and ibuprofen as antibacterial agents against red complex pathogens. *J Periodontol*. 2019;90(12):1441–8.
33. Paramasivam A, Vijayashree Priyadharsini J, Raghunandhakumar S. N6-adenosine methylation (m6A): a promising new molecular target in hypertension and cardiovascular diseases. *Hypertens Res*. 2020;43(2):153–4.
34. Vijayashree Priyadharsini J, Smiline Girija AS, Paramasivam A. An insight into the emergence of *Acinetobacter baumannii* as an oro-dental pathogen and its drug resistance gene profile - An in silico approach. *Heliyon*. 2018;4(12):e01051.
35. Paramasivam A, Vijayashree Priyadharsini J. Novel insights into m6A modification in circular RNA and implications for immunity. *Cell Mol Immunol*. 2020;17(6):668–9.
36. Paramasivam A, Priyadharsini JV, Raghunandhakumar S. Implications of m6A modification in autoimmune disorders. *Cell Mol Immunol*. 2020;17(5):550–1.
37. Girija ASS, Shankar EM, Larsson M. Could SARS-CoV-2-Induced Hyperinflammation Magnify the Severity of Coronavirus Disease (CoViD-19) Leading to Acute Respiratory Distress Syndrome? *Front Immunol*. 2020;11:1206.
38. Jayaseelan VP, Arumugam P. Exosomal microRNAs as a promising theragnostic tool for essential hypertension. *Hypertens Res*. 2020;43(1):74–5.
39. Ushanthika T, Smiline Girija AS, Paramasivam A, Priyadharsini JV. An in silico approach towards identification of virulence factors in red complex pathogens targeted by reserpine. *Nat Prod Res*. 2021; 35(11):1893–8.
40. Ramalingam AK, Selvi SGA, Jayaseelan VP. Targeting prolyl tripeptidyl peptidase from *Porphyromonas gingivalis* with the bioactive compounds from *Rosmarinus officinalis*. *Asian Biomed*. 2019;13(5):197–203.
41. Kumar SP, Girija ASS, Priyadharsini JV. Targeting NM23-H1-mediated inhibition of tumour metastasis in viral hepatitis with bioactive compounds from *Ganoderma lucidum*: A computational study. *pharmaceutical-sciences* [Internet]. 2020; 82(2). Available:<https://www.ijpsonline.com/article/s/targeting-nm23h1mediated-inhibition-of-tumour-metastasis-in-viral-hepatitis-with-bioactive-compounds-from-ganoderma-lucidum-a-comp-3883.html>
42. Mathivadani V, Smiline AS, Priyadharsini JV. Targeting Epstein-Barr virus nuclear antigen 1 (EBNA-1) with *Murraya koenigii* bio-compounds: An in-silico approach. *Acta Virol*. 2020;64(1):93–9.
43. Samuel SR, Kuduruthullah S, Khair AMB, Shayeb MA, Elkaseh A, Varma SR. Dental pain, parental SARS-CoV-2 fear and distress on quality of life of 2 to 6 year-old children during COVID-19. *Int J Paediatr Dent*. 2021;31(3):436–41.
44. Samuel SR. Can 5-year-olds sensibly self-report the impact of developmental enamel defects on their quality of life? *Int J Paediatr Dent*. 2021;31(2):285–6.
45. Barma MD, Muthupandiyan I, Samuel SR, Amaechi BT. Inhibition of *Streptococcus mutans*, antioxidant property and cytotoxicity of novel nano-zinc oxide varnish. *Arch Oral Biol*. 2021;126:105132.

46. Teja KV, Ramesh S. Is a filled lateral canal -A sign of superiority? J Dent Sci. 2020; 15(4):562–3.
47. Reddy P, Krithikadatta J, Srinivasan V, Raghu S, Velumurugan N. Dental Caries Profile and Associated Risk Factors Among Adolescent School Children in an Urban South-Indian City. Oral Health Prev Dent. 2020;18(1):379–86.
48. Jayaseelan VP, Paramasivam A. Emerging role of NET inhibitors in cardiovascular diseases. Hypertens Res. 2020;43(12): 1459–61.
49. Iswarya Jaisankar A, Smiline Girija AS, Gunasekaran S, Vijayashree Priyadharsini J. Molecular characterisation of csgA gene among ESBL strains of A. baumannii and targeting with essential oil compounds from Azadirachta indica. Journal of King Saud University - Science. 2020;32(8): 3380–7.
50. Girija AS. Fox3 (+) CD25 (+) CD4 (+) T-regulatory cells may transform the nCoV's final destiny to CNS! COMMENT. WILEY 111 River ST, Hoboken 07030-5774, NJ USA;2021.

© 2021 Saivarshine and Muralidharan; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:

<https://www.sdiarticle5.com/review-history/74346>