

International Journal of Pathogen Research

8(2): 1-5, 2021; Article no.IJPR.74873

ISSN: 2582-3876

Detection and Antibiogram Profile of *Enterococcus*Species from Local Cheese Sample within Ikare and Akungba Akoko, Ondo State Nigeria

Ayodeji Charles Osunla^{1*}

¹Department of Microbiology, Adekunle Ajasin University, Akungba-Akoko, P.M.B 001, Nigeria.

Author's contributions

The sole author designed, analyzed, interpreted and prepared the manuscript.

Article Information

DOI: 10.9734/IJPR/2021/v8i230198

Editor(s)

(1) Prof. John Yahya I. Elshimali, UCLA School of Medicine & Charles R. Drew University of Medicine and Science, USA. Reviewers:

(1) Simon Mburu Njoroge, Moi university, Kenya.

(2) Jaya Prakash, Sacred Heart College (Autonomous), India.

Complete Peer review History: https://www.sdiarticle4.com/review-history/74873

Original Research Article

Received 18 July 2021 Accepted 28 September 2021 Published 06 October 2021

ABSTRACT

Enterococci are part of the normal intestinal flora of humans and animals and are increasingly recognized as significant human pathogens and capable of causing major therapeutic challenges. The aim of this study was to isolate, identify and determine the antimicrobial susceptibility pattern of Enterococcus species in two local cheese sample collected from Akungba and Ikare. The isolation of Enterococcus was carried out using standard culture-based techniques. Antimicrobial susceptibility profile of the Enterococcus species was determined using disk diffusion method. A total number of 93 presumptive Enterococcus species were recovered from forty two different samples over a period of three months. The bacterial count observed on the Bile Aesculin agar ranged from 1.5x104 cfu/ml to 4.6x104 cfu/ml with Akungba sample having the highest bacterial load of 4.6x10⁴ cfu/ml. The Morphology and biochemical characteristics of suspected Enterococcus spp. isolated from the cheese sample revealed Enterococci feacalis as the isolated bacteria. The isolated Enterococcus species were tested against a panel of six antibiotics which include Penicillin G, Vancomycin, Tetracycline, Nitrofurantoin, Ciprofloxacin and Imipenem. It was observed that the isolates were susceptible to tetracycline, imipinem and ciprofloxacin while they are resistant to ciprofloxacin and vancomycin. The prevalence of Enterococci was confirmed with Enterococcus feacium and Enterococcus faecalis as the predominant species isolated in both cheese sample. The ability of *Enterococcus* species to survive a range of adverse environments allows multiple

^{*}Corresponding author: Email: osunlacharles@gmail.com;

routes of cross-contamination of *Enterococci* in causing human disease, including those from food. Overall, greater understanding of the ability of *Enterococcus* species to survive stresses, of virulence traits and especially of increasing antibiotic resistance, is needed in order to fully appreciate the complexity of *Enterococcus* species in causing disease.

Keywords: Enterococcus; cheese; antibiotics resistance; environments; disease.

1. INTRODUCTION

Enterococci are Gram-positive bacteria ubiquitously found in the gastrointestinal tracts of mammals, as well as in soil and water [1]. Food, especially when eaten raw, is a possible intermediate vehicle for the transmission of multidrug and/or virulent Enterococci strains from animal origin that can persist in the human intestinal tract [2-4]. Enterococci are mainly opportunistic pathogen that are harmless in healthy individuals and mainly cause infection in patients who are in intensive care units, with underlying disease or who are immunecompromised [5]. Enterococci are ubiquitous bacteria which colonize different niches. The primary habitat is considered to be gastrointestinal tract of animals and humans, thus via fecal contamination reach the raw milk and meat [6]. The wide distribution of the Enterococci, their resistance high temperatures, tolerance to high salt concentration augmented their survival through the production process and implication that this group of organisms, may also be isolated from heat treated milk and dairy products, especially cheese made from raw milk [7]. Enterococci, present in raw milk, may develop during cheese making process and ripening period and may represent the predominant microflora found in cheese made from raw milk [8]. It is noteworthy that Enterococcus species are also predominant pathogens causing bovine mastitis, which affects udder health and milk quality [9,10]. Consequently, contamination of raw cheeses with animal, human or environmental Enterococcal strains during different stages of processing is likely to occur. Of note, different studies investigating the microbiological quality of raw milk cheeses confirmed the difficulties in obtaining safe raw milk cheeses for consumption in many countries all over the world [11] Within genus Enterococcus, two species. Enterococcus and Enterococcus faecalis faecium, have emerged opportunistic pathogens and are responsible for an increasing percentage of nosocomial infections, including bacteremia and intraabdominal and urinary tract infections [12].

Recently, the evolution of hospital acquired multidrug-resistant *E. faecium* strains from *E. faecium* of animal origin has been elucidated [13] and indicates that *E. faecium* of animal origin can act as a donor of antibiotic resistance genes to other pathogenic *Enterococci*. The aim of this study was to carry out quality indices survey of *Enterococcus* spp. from the local cheese samples in Akungba Akoko, Ondo State, Nigeria.

2. MATERIALS AND METHODS

2.1 Study Area

Fresh and processed Wara and Nunu samples were collected from two different towns in Ondo State which includes; Akungba-Akoko and Ikare-Akoko.

2.2 Sample Collection

Samples were obtained from Akungba and Ikare between July and September, 2018. Wara and Nunu samples were obtained using 40ml sterile screw capped plastic bottles and immediately placed on ice. Samples were thereafter conveyed to the laboratory for microbiological analysis within 2 h after collection.

2.3 Enumeration and Isolation of Bacterial Isolates

An aliquot of 0.1 µl of 10-4 dilution of each sample was inoculated into the Nutrient agar (Oxoid, United Kingdom) and Bile Esculin Azide agar (Oxoid, United Kingdom) for heterotrophic and enterococci enumeration respectively. All Media were sterilized by autoclaving at 121°C for 15mins.After incubation, bacterial colonies were enumerated on the Nutrient agar and expressed as colony forming units per ml (cfu/ml). Black hallow colonies on Bile Esculin Azide agar were considered as presumptive enterococci colonies and were enumerated and expressed as cfu/ ml. Thereafter, an average of 3 to 4 enterococci isolates was selected per positive plate and subcultured on fresh Bile Esculin Azide agar and incubated aerobically at 37 °C for 18-24 h. Isolates were then purified on Nutrient agar and stored on agar slants at 4 °C until ready for further analysis.

3. RESULTS

The bacterial count observed on the Bile Aesculin agar ranged from 1.5×10^4 cfu/ml to 4.6×10^4 cfu/ml with Akungba sample having the highest bacterial load of 4.6×10^4 cfu/ml while Akungba sample had the lowest microbial load with 1.5×10^4 cfu/ml. The Morphology and biochemical characteristics of suspected Enterococcus spp. isolated from the cheese sample is shown in Table 1 which reveal *Enterococci feacalis* and *Enterococcus faecium* as the isolated bacteria. The biochemical tests

such as oxidase test and sugar tests such as lactose, glucose, maltose, mannitol, sucrose and fructose carried out and shown in the table also supported the identification of the presumptive isolates as listed in the table

The isolated Enterococcus spp were tested against a panel of six antibiotics and the results are tabulated based on the cattle farm in which the study was carried out. The antibiotics used were Penicillin G, Vancomycin, Tetracycline, Nitrofurantoin, Ciprofloxacin and Imipenem. It was observed that the isolates were susceptible to Tetracycline, Imipinem and Ciprofloxacin but they are resistant to Penicillin G, ciprofloxacin and vancomycin.

Table 1. Morphology and biochemical characteristics of representative presumptive Enterococcus species recovered from cheese samples

Isolates	Shape	Oxidase	Mannitol	Glucose	Sucrose	Fructose	Lactose	Probable identity
1	Cocci	_	Ag	Ag	Ag	Ag	Ag	Enterococcus feacium
2	Cocci	_	Ag	Ag	Ag	Ag	Ag	Enterococcus faecium
3	Cocci	_	Ag	Ag	Ag	Ag	Ag	Enterococcus feacalis
4	Cocci	_	Ag	Ag	Ag	Ag	Ag	Enterococcus feacium
5	Cocci	_	Ag	Ag	Ag	Ag	Ag	Enterococcus feacalis

Key: = Negative; Ag = Acid gas positive

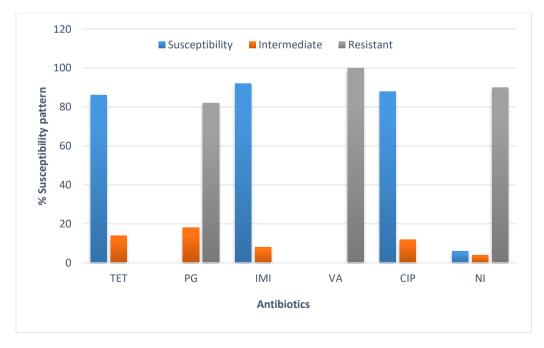


Fig. 1. Antibiotic susceptibility profiles of presumptive *Enterococcus* species isolated from selected rivers

Key: TET = Tetracycline; PG = Penicillin G; IMI = Imipenem; CIP = Ciprofloxacin; Ni = Nitrofurantoin

4. DISCUSSION

The genus Enterococcus is a diverse group of bacteria that have been reported in a variety of niches with gastrointestinal tract being their principal habitat [14]. The prevalence Enterococci was confirmed with Enterococcus feacium and Enterococcus faecalis biochemically as the predominant species isolated in both cheese sample. The identification Enterococcus species is vital because for many years Enterococci were believed to be harmless humans and considered unimportant medically. Recently. Enterococci have become one of the most common nosocomial pathogens, giving a high mortality rate of up to 61 % [15]. The ability of *Enterococcus* species to survive a range of adverse environments allows multiple routes of cross-contamination of Enterococci in causing human disease, including those from food. The occurrence of resistant and virulent strains of Enterococcus species in farm animals could be detrimental to both the animals and the population. The high counts Enterococcus species obtained indicated that the cheese samples might have been contaminated by either the anthropogenic activities of the food handlers or the cattle is infected. This study provides antibiotic information about the resistance patterns of Enterococci isolated from cattle farms as well as their virulent potentials. Previous studies have shown that the occurrence of resistant strains of Enteroccoccus in cattle farms is due to the overuse of antibiotics in farm management [16]. The increasing concern about this organism is their occurrences and the spread antibiotic resistance genes (Lester et al., 2011). The isolated strains of Enterococci were resistant to following antibiotics: penicillin G, vancomycin and nitrofurantoin. Bulajić and Mijačević [17] reported forty two strains of isolated Enterococci, which were subjected to antibiotic susceptibility testing. They reported that the isolated strains of Enterococci were resistant to the following antibiotics: penicillin, tetracycline, lincomycin, gentamycin, neomycin, erythromycin and chloramphenicol. In another study that was carried out in Thailand, a significant prevalence of MDR enterococci (10.3%) were Vancomycin resistant Enterococcus (VRE) isolates, in water samples including agricultural wells of food animal farms, canals and rivers [18]. Antibiotic resistance could be associated to the abuse of antibiotics due to over-the-counter medication and accessibility to patients devoid of doctor's prescription due to upsurge of pharmacies and patent medicine stores. In addition, consumption

of animal products treated with these antibiotics as growth promoters and prophylactic agents could have added to the resistance [19]. In order evaluate the potential risk of contaminated with resistant strains Enterococci represent to human health, it is important to distinguish between intrinsic and acquired antibiotic resistance [20,21]. Further investigations are needed to address this question.

5. CONCLUSION

Overall, greater understanding of the ability of *Enterococcus* species to survive stresses, of virulence traits and especially of increasing antibiotic resistance, is needed in order to fully appreciate the complexity of *Enterococcus* species in causing disease.

CONSENT

As per international standard or university standard, patients' written consent has been collected and preserved by the author(s).

ETHICAL APPROVAL

As per international standard or university standard written ethical approval has been collected and preserved by the author(s).

COMPETING INTERESTS

Author has declared that no competing interests exist.

REFERENCES

- Castillo-Rojas G, Mazari-Hiriart M, de-Leon PS, Amieva-Fernandez RI, Agis-Juarez RA. Comparison of Enterococcus faecium and Enterococcus faecalis strains isolated from water and clinical samples: Antimicrobial susceptibility and genetic relationships. PLoS ONE. 2013;8(4): 59481-59491.
- 2. Jamet E, Akary E, Poisson M-A, Chamba J-F, Bertrand X, Serror P. Prevalence and characterization of antibiotic resistant *Enterococcus* faecalis in French cheeses. Food Microbiology. 2012;31:191e198.
- Kayser F. Safety aspects of Enterococcifrom the medical point of view. International Journal of Food Microbiology. 2003;88:255e262.

- 4. Novais C, Coque T, Costa M, Sousa J, Baquero F, Peixe L. High occurrence and persistence of antibiotic-resistant Enterococciin poultry food samples in Portugal. Journal of Antimicrobial Chemotherapy. 2005; 56:1139e1143.
- 5. Martins E, Novais N, Freitas AR, Dias AR, Ribeiro TG, Antunes P, Peixe L. Filling the map for antimicrobialresistance in Sub-Saharan Africa:Ampicillin-resistant *Enterococcus* fromnon-clinical sources in Angola. Journal of Antimicrobial Chemotherapy. 2015;70:42-51.
- Mariam SH. A sampling survey of Enterococciwithin pasteurized, fermented dairy products and their virulence and antibiotic resistance properties. Plos one. 2021;16(7):e0254390.
- 7. Dapkevicius MDLE, Sgardioli B, Câmara S, Poeta P, Malcata FX. Current Trends of Enterococci in Dairy Products: A Comprehensive Review of Their Multiple Roles. Foods. 2021; 10(4):821.
- 8. Fortina MG, Ricci G, Borgo F, Manachini PL, Arends K, Schiwon K, Abajy MY, Grohmann E. A survey on biotechnological potential and safety of the novel *Enterococcus* species of dairy origin, E. italicus. International journal of food microbiology, 2008;123(3):204-211.
- Rysanek D, Zouharova M, Babak V. Monitoring major mastitis pathogens at the population level based on examination of bulk tank milk samples. Journal of Dairy Research, 2009;76(1):117-123.
- Rysanek D, Zouharova M, Babak V. Monitoring major mastitis pathogens at the population level based on examination of bulk tank milk samples. J Dairy Res. 2009;76:117-123.
- Little C, Rhoades J, Sagoo S, Harris J, Greenwood M, Mithani V, et al. Microbiological quality of retail cheeses made from raw, thermized or pasteurized milk in the UK. Food Microbiology. 2008;25:304e312.
- Conde-Estevez D, Grau S, Albanell J, Terradas R, Salvado M, Knobel H. Clinical characteristics and outcomes of patients with vancomycin-susceptible *Enterococcus*

- faecalis and *Enterococcus* faecium bacteraemia in cancer patients. European Journal of Clinical Microbiology and Infectious Diseases. 2011;30:103-108.
- Lebreton F, van Schaik W, Mc Guire AM, Godfrey P, Griggs A, Mazumdar V. Emergence of epidemic multidrug-resistant Enterococcus faecium from animal and commensal strains. MBio. 2013;4:e00534-00513.
- Lebreton F, Willems RJ, Gilmore MS. *Enterococcus* diversity, origins in nature, and gut colonization. Enterococci: from commensals to leading causes of drug resistant infection; 2014.
- Buetti N, Wassilew N, Rion V, Senn L, Gardiol C, Widmer A, Marschall J. Emergence of vancomycin-resistant Enterococciin Switzerland: a nation-wide survey. Antimicrobial Resistance & Infection Control. 2019;8(1):1-5.
- Phares CA, Danquah A, Atiah K, Agyei FK, Michael OT. Antibiotics utilization and farmers' knowledge of its effects on soil ecosystem in the coastal drylands of Ghana. PloS one. 2020; 15(2):e0228777.
- 17. Bulajić S, MijačevićZ. Enterococci in cheese phenotypization and antibiotic resistance. Acta agriculturae slovenica. 2004;84(1):25–30.
- Tansuphasiri U, Khaminthakul D, Pandii W. Antibiotic resistance of Enterococciisolated from frozen foods andenvironmental water. Southeast AsianJournal of Tropical Medicine and PublicHealth. 2006;37:162–170.
- Beshiru A, Igbinosa IH, Omeje FI, Ogofure AG, Eyong MM, Igbinosa EO. Multiantibiotic resistant andputative virulence gene signatures in *Enterococcus* species isolated from pig farms environment. MicrobialPathogenesis. 2017;104:90-96.
- 20. Clewell DB. Movable genetic elements and antibiotic resistance in Enterococci. European Journal of Clinical Microbiological Infection Diseases. 1990;9:90–102.
- 21. Murray BE. The life and times of *Enterococcus*. Clinical Microbiological Reviews. 1990;3:46–65.

© 2021 Osunla; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:
The peer review history for this paper can be accessed here:
https://www.sdiarticle4.com/review-history/74873