

Journal of Scientific Research and Reports

Volume 30, Issue 10, Page 509-515, 2024; Article no.JSRR.117976 ISSN: 2320-0227

Effect of Different Kharif Paddy Straw Management Options and Different Levels of Nitrogen on Soil Dehydrogenase and Urease Activity

Navya. K a*, Uma Reddy. R ^a , Krishna Chaitanya. A ^a and Suneetha Devi. K. B ^b

^a Department of Soil Science and Agricultural Chemistry, Agricultural College, Professor Jayashankar Telangana State Agricultural University, Polasa, Jagtial, India. ^b Department of Agronomy, Agricultural College, Professor Jayashankar Telangana State Agricultural University, Polasa, Jagtial, India.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI:<https://doi.org/10.9734/jsrr/2024/v30i102478>

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/117976>

Original Research Article

Received: 01/08/2024 Accepted: 03/10/2024 Published: 07/10/2024

ABSTRACT

A field experiment was conducted during *rabi* 2020-21**,** at Regional Agricultural Research Station, Polasa, Jagtial, PJTSAU to study soil dehydrogenase and urease activity as influenced by different *kharif* paddy straw management options and nitrogen levels. The results reveal that dehydrogenase and urease activity of soil was significantly influenced by paddy straw management options and activity was increased with crop duration up to 60 DAT and decreased after harvest. Dehydrogenase activity was reduced after burning (1.93 mg TPF produced g⁻¹ soil d⁻¹) as compared to the initial values (2.34 mg TPF produced g^{-1} soil d⁻¹). The highest dehydrogenase activity of 3.74 and 3.54 mg TPF produced g^{-1} soil d⁻¹ was recorded at 60 DAT in paddy straw incorporation with

^{}Corresponding author: Email: navyakaringula49@gmail.com;*

Cite as: K, Navya., Uma Reddy. R, Krishna Chaitanya. A, and Suneetha Devi. K. B. 2024. "Effect of Different Kharif Paddy Straw Management Options and Different Levels of Nitrogen on Soil Dehydrogenase and Urease Activity". Journal of Scientific Research and Reports 30 (10):509-15. https://doi.org/10.9734/jsrr/2024/v30i102478.

phosphorus and incorporation without phosphorus and decreased to 3.36 and 3.03 mg TPF produced g-1 soil d-1 after harvest respectively. Soil urease activity was showed an increasing trend with crop duration up to 60 DAT and then declined. Maximum activity of 3.05, 4.84, 5.17 mg NH₄+ released g^{-1} soil h⁻¹ was observed at 60 DAT under paddy straw burning, paddy straw incorporation with phosphorus incorporation without phosphorus respectively. Paddy straw incorporation with phosphorus recorded higher urease activity and paddy straw burning recorded lower urease activity at all the intervals of crop growth. The effect of nitrogen levels on soil urease activity was found to be significant. Soil urease was highest with 120% RDN followed by 115% RDN, followed by 110% RDN, followed by 100% RDN and the similar trend was observed at all the intervals of crop growth.

Keywords: Paddy straw; burning; incorporation; nitrogen.

1. INTRODUCTION

Rice is the most residue producing crop. In future with increase in grain production will led to enormous generation of straw. In India, straw is produced in about 168.50 million tonnes [1]. The intensification of the cropping system resulting in production of paddy straw in large volumes which need to be managed over shorter turnaround period between the crops. Globally, roughly 800 to 1000 Mt per annum of rice straw is produced, with about 600 to 800 Mt per annum produced in Asia (Bhuvaneshwari et al., 2019). And due to scarcity of labour and high labour cost involved in its removal from the field and also with little management options burning of paddy straw is adapted and it has been increased dramatically over the last decade.

One of the advantages of burning is that it is cost effective and the land will be cleared quickly from residues before the next crop is taken up, thus facilitating establishment of next succeeding crop. Though it has some positive effects in managing pests, it also leads to loss of essential soil nutrients – upto 80 % N, 25% P, 21% K and 4-60% S and 13 tonnes per ha of carbondioxide causing loss of soil organic matter [2], reduces microbial diversity [3].

Several researchers advocated paddy straw incorporation in surface soil as an alternative to rice straw burning, saying that it improves soil quality and increases SOC by giving energy and substrates to microbes through soil enzyme activities [4-6].

The purpose of this research was to see how N application and paddy straw integration affected soil dehydrogenase and urease activities.

2. MATERIALS AND METHODS

The experiment was conducted at Regional agricultural research station, Polasa, Jagtial district. It is situated in the Northern Telangana Zone. The experimental site is located between 18º 51' 53'' N latitude and 78º 56'21'' E longitude at an altitude of 243.4 m above mean sea level (MSL). The soil of the experimental site was sandy clay loam, which is medium in organic carbon (0.54%), low in soil available nitrogen 157.6 kg ha-1 , high in soil available phosphorus $(31.05kg$ ha⁻¹) and potassium $(309.5kg$ ha⁻¹). The experiment was laid out in Factorial RBD, with 2 factors, factor 1 comprised of 3 levels and factor 2 comprised of 4 levels with a total of 12 treatments and replicated thrice. Paddy straw mulcher was run to chop it and is incorporated 10 days before transplanting. Paddy (variety- JGL-24423) was transplanted with a spacing of 15x15cm. The recommended dose of fertilizer was 150:60:40 kg N, P_2O_5 , K₂O ha-1 . The entire recommended dose of phosphorus and potassium was applied as basal dose in the form of SSP and MOP respectively. Nitrogen was applied in 3 splits and the excess of RDN was applied at basal in the form of urea.

2.1 Dehydrogenase Activity

A 1 g of soil sample taken in an air tight screw capped test tubes and it was saturated with 0.2 ml of 3% triphenyl tetrazolium chloride solution. 0.5 ml of 1% glucose solution was added to the tubes and the bottom of the tubes are tapped to remove the entrapped air and a water seal was formed above the soil. The tubes were incubated for 24 h at 28±0.5°C. 10 ml of methanol was added to the incubated samples and shaken vigorously and were allowed to stand for 6 h. Clear pink coloured supernatant liquid was withdrawn and readings were taken with spectrophotometer at a wavelength of 485 nm. The results were expressed as mg of triphenyl formazan (TPF) g-1 d -1 [7].

List 1. Treatment details

- T_1 100% RDN + Residue burning
- T_2 10% excess N over RDN + Residue burning
 T_3 15% excess N over RDN + Residue burning
- 15% excess N over RDN + Residue burning
- T⁴ 20% excess N over RDN + Residue burning
- T_5 100% RDN + Residue incorporation without phosphorus
- T_6 10% excess N over RDN + Residue incorporation without phosphorus
- T_7 15% excess N over RDN + Residue incorporation without phosphorus
- T_8 20% excess N over RDN + Residue incorporation without phosphorus
- $T₉$ 100% RDN + Residue incorporation with P application through SSP
- T¹⁰ 10% excess N over RDN + Residue incorporation with P application through SSP
- T_{11} 15% excess N over RDN + Residue incorporation with P application through SSP
- T₁₂ 20% excess N over RDN + Residue incorporation with P application through SSP

2.2 Urease Activity

This method is based on determination of NH³ released other incubation of soil with urea solution for 2 hours at 300℃ [8]. To a 5 g of soil was taken in duplicate in 50 ml volumetric flask. 0.2 ml toluene and 9 ml of THAM buffer (pH- 9; 0.05M) were added to it. The flasks were swirled for few second to mix the content. Then 1 ml of 0.2 M urea solution was added and swirled again for a few second. The flasks were then stoppered and placed in an incubator at 300 ̊C for 2 h. After 2 h the stoppers were removed and approximately 35 ml of $KCI - Ag₂SO₄$ solution was added and the flasks were swirled for few seconds and allowed to stand until the contents have cooled to room temperature. The volume of the flasks was made upto mark (50 ml) by addition of $KCI - Ag₂SO₄$ solution. The flasks were stoppered and inverted several times to mix the contents. To perform control, the above procedure was followed, but 1 ml of 0.2M urea solution was added after the addition of 35 ml KCl- Ag2SO⁴ solution.

A 40 ml aliquot of the suspension was pipetted out into 100 ml distillation flask and 0.2 g MgO was added to it for determination of NH₄+-N in the resulting soil suspension. The content of the flask was then distilled for 15 minutes and the distillate was collected in a 50 ml conical flask containing 5 ml of 2 % boric acid indicator. The distillate was then titrated with 0.005 *N*sulphuric acid. The urease activity is expressed as mg NH_4 +-N g⁻¹ soil⁻¹ h⁻¹ at 300 °C.

3. RESULTS AND DISCUSSION

3.1 Dehydrogenase Activity (mg TPF Produced g-1 Soil d-1)

Activity of dehydrogenase reflects oxidative activity of soil microflora and is a good indicator of microbial activity and the greater amount of soil organic carbon stimulates dehydrogenase activity [9]. The changes in soil dehydrogenase activity (mg TPF produced g^{-1} soil d⁻¹) at different crop growth intervals are furnished in Table 1.

The results revealed that dehydrogenase activity of soil was significantly influenced by paddy straw management and activity was increased with crop duration up to 60 DAT and decreased after harvest. Dehydrogenase activity was reduced after burning (1.93 mg TPF produced g-1 soil d^{-1}) as compared to the initial values (2.34 mg TPF produced g^{-1} soil d⁻¹). The results were in same line with Kumar et al. [10]. Decrease in soil dehydrogenase activity in burnt treatments indicates lower microbial activity. Burning of crop residue increases soil temperature [11]. which kills bacterial and fungal population. About 50% of bacterial population was killed by burning paddy straw [12] and loss of soil carbon from the soil [13]. However, its activity was regained at later stages [14]. Dehydrogenase showed an increasing trend with time. The Dehydrogenase activity was higher in the treatment with paddy straw incorporation with phosphorus followed by paddy straw incorporation without phosphorus and lower in paddy straw burning treatment. The highest dehydrogenase activity of 3.74 mg TPF produced g^{-1} soil d⁻¹ was recorded at 60 DAT in paddy straw incorporation with phosphorus and decreased to 3.36 mg TPF produced g^{-1} soil d^{-1} after harvest. While in case of paddy straw incorporation without phosphorus the highest dehydrogenase activity of 3.54 mg TPF produced g⁻¹ soil d⁻¹ was recorded at 60 DAT in paddy straw incorporation with phosphorus and decreased to 3.03 mg TPF produced g^{-1} soil d⁻¹ after harvest. Higher dehydrogenase activity in paddy straw incorporated treatments might be due to higher microbial activities during active decomposition of paddy straw [10] and increased

Fig. 1. Soil dehydrogenase(mg TPF produced g-1 soil d-1) (left), and urease activity (mg NH⁴ + released g-1 soil h-1)as influenced by paddy straw managementoptions

Fig. 2. Soil urease activity as influenced by different levels of nitrogen (mg NH⁴ + released g-1 soil h-1)

root activity. Crop residue incorporation to the soil adds large amount of labile organic carbon to the soil which then enhances soil dehydrogenase activity [15]. Soil becomes dry as the crop reaches maturity affecting microbial activity and thus the enzymatic activity [16].

The interaction between paddy straw management and nitrogen levels was found to be non-significant for dehydrogenase activity.

3.2 Urease Activity (mg NH⁴ ⁺ Released g-1 Soil h-1)

The data on soil urease activity (mg NH_4 + released g^{-1} soil h⁻¹) at different intervals is

furnished in Table 2. Urease enzyme catalyzes the hydrolysis of urea hydroxycarbamide and semi-carbazide to carbon dioxide and ammonia and plays an important role in nitrogen cycling [17]. Paddy straw management and nitrogen levels were found to influence the soil available nitrogen.

Soil urease activity has shown an increasing trend with crop duration up to 60 DAT and then declined. At 15 DAT, minimum activity of 1.97, 3.83, 4.44 mg NH₄⁺ released g^{-1} soil h⁻¹ was observed in paddy straw burning, paddy straw incorporation with phosphorus and paddy straw incorporation with phosphorus respectively. Maximum activity of 3.05, 4.84, 5.17 mg NH4⁺

Treatments	15 DAT	30 DAT	45 DAT	60 DAT	75 DAT	After Harvest				
Paddy straw management										
RB	2.01	2.13	2.40	2.59	2.33	2.23				
$I - P$	2.87	3.01	3.38	3.54	3.16	3.03				
$H + P$	3.17	3.28	3.59	3.74	3.47	3.36				
$SEm+$	0.07	0.08	0.08	0.10	0.12	0.09				
CD@ 5%	0.20	0.24	0.23	0.28	0.35	0.27				
Nitrogen levels										
100% RDN	2.51	2.64	2.97	3.12	2.84	2.70				
10% excess RDN	2.63	2.73	3.10	3.27	2.91	2.78				
15% excess RDN	2.71	2.83	3.15	3.32	3.03	2.94				
20% excess RDN	2.88	2.92	3.27	3.46	3.17	3.06				
SE _m	0.08	0.09	0.09	0.11	0.14	0.11				
CD@ 5%	ΝS	NS.	NS	ΝS	NS	NS.				
Interactions										
SE _m	0.14	0.16	0.16	0.19	0.24	0.19				
CD @ 5%	ΝS	NS.	NS	ΝS	NS.	NS.				
CV	8.88	10.29	8.87	10.14	13.71	11.25				

Table 1. Effect of different *kharif* **paddy straw management options and fertilizer N levels on soil dehydrogenase activity (mg TPF produced g-1 soil d-1) at different intervals of** *rabi* **rice**

Table 2. Effect of different *kharif* **paddy straw management options and fertilizer N levels on soil urease activity (mg NH⁴ + released g-1 soil h-1) at different intervals of** *rabi* **rice**

Treatments	15 DAT	30 DAT	45 DAT	60 DAT	75 DAT	After Harvest			
Paddy straw management									
RB	1.97	2.48	2.86	3.05	2.57	2.34			
I-P	3.83	4.33	4.61	4.84	4.31	4.12			
$H + P$	4.44	4.79	4.96	5.17	4.66	4.38			
SEm _±	0.08	0.10	0.10	0.11	0.10	0.09			
CD@ 5%	0.24	0.28	0.29	0.33	0.28	0.26			
Nitrogen levels									
100% RDN	3.03	3.61	3.83	4.00	3.52	3.35			
110% RDN	3.21	3.75	4.08	4.30	3.72	3.49			
115% RDN	3.57	3.94	4.27	4.42	4.01	3.77			
120% RDN	3.83	4.15	4.40	4.69	4.13	3.85			
SEm _±	0.09	0.11	0.11	0.13	0.11	0.10			
CD@ 5%	0.28	0.32	0.33	0.38	0.33	0.29			
Interactions									
SE _m	0.16	0.19	0.20	0.23	0.19	0.17			
CD @ 5%	ΝS	NS	ΝS	NS.	ΝS	NS.			
CV	8.30	8.57	8.18	9.03	8.73	8.33			
	PR-Residue Burning L-P-Incorporation with Phosphorus L-P-Incorporation with Phosphorus PDN-Recommended Dose of Nitrogen AH-At								

RB-Residue Burning, I-P-Incorporation with Phosphorus, I+P-Incorporation with Phosphorus, RDN-Recommended Dose of Nitrogen, AH-At Harvest The interaction between paddy straw management and nitrogen levels was found to be non significant.

released g^{-1} soil h⁻¹ was observed at 60 DAT under paddy straw burning, paddy straw incorporation with phosphorus and paddy straw incorporation with phosphorus respectively. Paddy straw incorporation with phosphorus recorded higher urease activity and paddy straw burning recorded lower urease activity at all the intervals of crop growth. When paddy straw added to soil, microbes immobilizes the mineral nitrogen which was later mineralized [18] indicating that during decomposition a portion of straw N could be transformed into mineral in soil nitrogen cycling (Bradford and Peterson, 2000). In addition urease transforms the soil mineral N into source of N available for

crop use at its later stages, which improves efficiency [19].

The effect of nitrogen levels on soil urease activity was found to be significant. Soil urease was the highest with 120% RDN followed by 115% RDN >110% RDN >100% RDN and the similar trend was observed at all the intervals of crop growth. The increase in soil urease activity with paddy straw management and nitrogen levels is in conformity with Sharma et al. [20], Singh and Sharma [21-23]. The increases might be due to straw addition enhances metabolic activity of microorganisms, microbial population thereby improving soil enzymatic activity [24-26] and Sharma et al., 2020a).

4. CONCLUSION

Soil dehydrogenase and urease activity was significantly influenced by paddy straw management. Dehydrogenase activity was reduced after burning. The highest dehydrogenase and urease was recorded at 60
DAT in paddy straw incorporation with straw incorporation with phosphorus and without phosphorus and
decreased after harvest. Paddy straw decreased after harvest. Paddy incorporation with phosphorus recorded higher dehydrogenase and urease activity and paddy straw burning recorded lower activity at all the intervals of crop growth. The effect of nitrogen levels on soil urease activity was found to be significant. Soil urease was highest with 120% RDN followed by 115% RDN, followed by 110% RDN, followed by 100% RDN and the similar trend was observed at all the intervals of crop growth.

DISCLAIMER (ARTIFICIAL INTELLIGENCE

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of this manuscript.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. FAO [Food and Agriculture Organization]. Statistical Database; 2017. Available: http://www.fao.org/statistics/en., 2017

[Accessed On:21 September 2020].

- 2. Mandal KG, Misra AK, Hati KM, Bandyopadhyay KK, Ghosh PK, Mohanty M. Rice residue-management options and effects on soil properties and crop productivity. Journal of Food Agriculture and Environment. 2004;2:224-231.
- 3. Zhang P, Wei T, Jia Z, Han Q, Ren X. Soil aggregate and crop yield changes with different rates of straw incorporation in semiarid areas of northwest China. Geoderma. 2014;230:41-49.
- 4. Bera T, Sharma S, Thind H.S, Yadvinder S, Sidhu HS, Jat ML. Soil biochemical changes at different wheat growth stages in response to conservation agriculture practices in a rice-wheat system of northwestern India*.* Soil Research. 2017;56:91– 104
- 5. Bera T, Sharma S, Thind HS, Yadvinder S, Sidhu HS, Jat ML. Changes in soil biochemical indicators at different wheat growth stages under conservation based sustainable intensification agriculture practices of rice-wheat system. Journal of Integrative Agriculture2018; 17:1871–1880
- 6. Saikia R, Sharma S, Thind HS, Sidhu HS, Yadvinder-Singh. Temporal changes in biochemical indicators of soil quality in response to tillage, crop residue and green manure management in a rice-wheat system. Ecological Indicators 2019; 103:383–394
- 7. Novak B. On the relation between dehydrogenase activity and $CO₂$ evolution in soil; 1972.
- 8. Tabatabai MA, Bremner JM. Assay of urease activity in soils. Soil Biology and Biochemistry. 1972a;4:479-487.
- 9. Velmourougane K, Venugopalan MV, Bhattacharyya T, Sarkar D, Pal DK, Sahu A, Ray SK, Nair KM, Prasad J, Singh RS. Soil dehydrogenase activity in agroecological sub regions of black soil regions in India. *Geoderma*. 2013;197-198:186- 192.
- 10. Kumar A, Kushwaha KK, Singh S, Shivay YS, Meena MC, Nain L. Effect of paddy straw burning on soil microbial dynamics in sandy loam soil of Indo-Gangetic plains. Environmental Technology and Innovation. 2019;16:100469.
- 11. Gupta PK, Sahai S, Singh N, Dixit CK, Singh DP, Sharma C, Tiwari MK, Gupta RK, Garg SC. Residue burning in rice– wheat cropping system: Causes and implications. Current Science. 2004;1713- 1717.
- 12. Hesammi E, Talebi AB, Hesammi A. A review on the burning of crop residue on the soil properties. WALIA Journal. 2014;30:192-194.
- 13. Mendham DS, O'connell AM, Grove TS, Rance SJ. Residue management effects on soil carbon and nutrient contents and growth of second rotation eucalypts. Forest Ecology and Management. 2003;181(3): 357-372.
- 14. Lohan SK, Jat HS, Yadav AK, Sidhu HS, Jat ML, Choudhary M, Peter JK, Sharma, PC. Burning issues of paddy residue management in north-west states of India. Renewable and Sustainable Energy Reviews. 2018;81:693-706.
- 15. Fang XM, Zhang XL, Chen FS, Zong YY, Bu WS, Wan SZ, Luo Y, Wang H. Phosphorus addition alters the response of soil organic carbon decomposition to nitrogen deposition in a subtropical forest. Soil Biology and Biochemistry. 2019;133:119-128.
- 16. Geisseler D, Horwath WR, Scow KM. Soil moisture and plant residue addition interact in their effect on extracellular enzyme

activity. Pedobiologia. 2011;54(2):71-78.

- 17. Sardans J, Penuelas J, Estiarte M. Changes in soil enzymes related to C and N cycle and in soil C and N content under prolonged warming and drought in a Mediterranean shrubland. Applied Soil Ecology. 2008;39(2):223-235.
- 18. Witt C, Cassman KG, Olk DC, Biker U, Liboon SP, Samson MI, Ottow JCG. Crop rotation and residue management effects on carbon sequestration, nitrogen cycling and productivity of irrigated rice systems. Plant and Soil. 225(1):263-278.
- 19. Zibilske LM, Bradford JM, Smart JR. Conservation tillage induced changes in organic carbon, total nitrogen and available phosphorus in a semi-arid alkaline subtropical soil. Soil and Tillage Research. 2002;66(2):153-163.
- 20. Sharma S, Singh P, Choudhary OP. Nitrogen and rice straw incorporation impact nitrogen use efficiency, soil nitrogen pools and enzyme activity in rice-wheat system in north-western India. Field Crops Research. 2000;266:108131.
- 21. Sharma S, Singh P, Kumar S. Responses of soil carbon pools, enzymatic activity,

and crop yields to nitrogen and straw incorporation in a rice-wheat cropping system in north-western India. Frontiers in Sustainable Food Systems. 2020;4: 203.

- 22. Singh RK, Sharma GK, Kumar P, Singh SK, Singh R. Effect of crop residues management on soil properties and crop productivity of rice-wheat system in inceptisolsof Seemanchal region of Bihar. Current Journal of Applied Science and Technology. 2019;37(6):1-6.
- 23. Navya K, Uma Reddy R, Krishna Chaitanya A, Suneetha Devi KB. Effect of different kharif paddy straw management options and nitrogen levels on soil organic carbon and soil total carbon and nitrogen. Int. J. Plant Soil Sci. 2024;36(5):763-70. [Accessed On:2024 May 22] Available:https://journalijpss.com/index.ph p/IJPSS/article/view/4574
- 24. Jiao XG, Gao CS, Lü GH, Sui YY. Effect of long-term fertilization on soil enzyme activities under different hydrothermal conditions in Northeast China. Agricultural Sciences in China. 2011;10:412–422.
- 25. Mahapatra SS, Parameswaran C, Chowdhury T, Senapati A, Chatterjee S, Singh AK, Panneerselvam P. Unraveling the Efficient Cellulolytic and Lytic Polysaccharide Monooxygenases Producing Microbes from Paddy Soil for Efficient Cellulose Degradation. J. Adv. Biol. Biotechnol. [Internet]. 2024;27(3):47- 56.

[Accessed on:2024 May 22] Available:https://journaljabb.com/index.php /JABB/article/view/720

26. Li XZ, Han BS, Yang F, Hu CY, Han GZ, Huang LM. Effects of land use change on soil carbon and nitrogen in purple paddy soil. Journal of Environmental Management. 2022;314: 115122.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of the publisher and/or the editor(s). This publisher and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.

___ *© Copyright (2024): Author(s). The licensee is the journal publisher. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.*

> *Peer-review history: The peer review history for this paper can be accessed here: <https://www.sdiarticle5.com/review-history/117976>*